

CMRC High-Speed Cell Sorting Facility

Experiment Information Sheet

User: _____ Date: _____

E-mail: _____ Phone: _____

PI: _____ Cost Center #: _____

Experimental Goal of Sort:

Cells to be Sorted:

(Please use one form for each sample or type of cells to be sorted, unless samples are similar and have same sorting parameters)

Cell type: cell line primary cells other _____

Species: human mouse other _____

Tissue origin: bone marrow peripheral blood neural other _____

Adhesiveness: non-adherent cell line adherent cell line

non-adherent tissue disaggregated tissue

Mode of disaggregation trituration trypsin collagenase other _____

Estimated cell size: small (~10 μ m, e.g., lymphocytes) large

Number of starting cells (#):

Volume of starting cells (ml):

Concentration of starting cells (#/ml):

Carrier solution: PBS HEPES Hanks BSA ___% FCS ___% other _____

Biohazard Consideration:

Are there exogenous genes transferred into the cells to be sorted: No Yes

If "Yes", list the genes here: _____

Are any of these genes oncogenes or pathogenic genes? No Yes

Method of gene transfer:

Chemical or physical method (Ca phosphate, lipofusion, electroporation)

Viral vector: Adenovirus Retrovirus Lentivirus

Name of vector: _____ Source of vector: _____

Name of packaging cell line: _____

Laboratory where the virus was prepared: _____

The viral preparation has been shown to be free of replication-competent virus:

Yes No

Sorting Parameters:

(Common fluorochromes/fluorescent proteins are shown in bold. For PI, BrdU, other DNA/RNA dyes, please consult.)

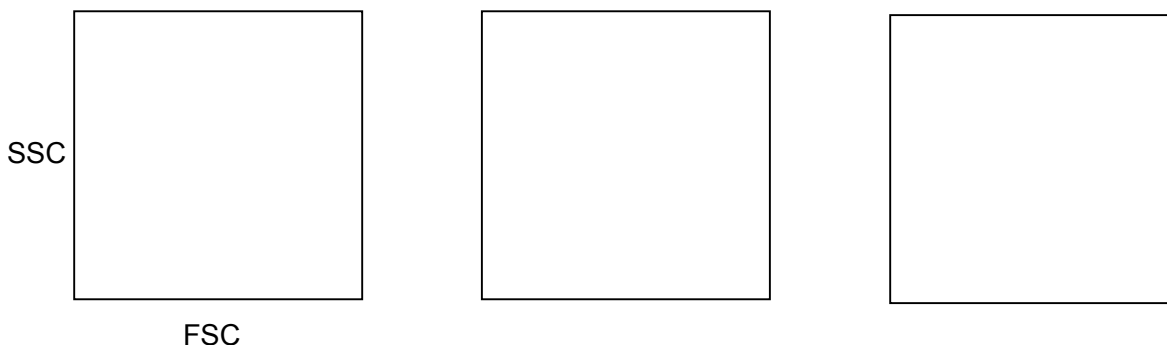
<input type="checkbox"/> FITC/GFP	(519 nm)	Antigen/markers: _____
<input type="checkbox"/> PE/DsRed	(578 nm)	Antigen/markers: _____
<input type="checkbox"/> PE-Texas Red	(610 nm)	Antigen/markers: _____
<input type="checkbox"/> PE-Cy5/PerCP-Cy5.5	(695 nm)	Antigen/markers: _____
<input type="checkbox"/> PE-Cy7	(785 nm)	Antigen/markers: _____
<input type="checkbox"/> APC/HcRed	(660 nm)	Antigen/markers: _____
<input type="checkbox"/> APC-Cy7	(785 nm)	Antigen/markers: _____
<input type="checkbox"/> DAPI	(450 nm)	Antigen/markers: _____
<input type="checkbox"/> AlexaFluor 430	(530 nm)	Antigen/markers: _____

Single-stained Controls Provided by User:

<input type="checkbox"/> Unstained	
<input type="checkbox"/> FITC/GFP	(519 nm)
<input type="checkbox"/> PE/DsRed	(578 nm)
<input type="checkbox"/> PE-Texas Red	(610 nm)
<input type="checkbox"/> PE-Cy5/PerCP-Cy5.5	(695 nm)
<input type="checkbox"/> PE-Cy7	(785 nm)
<input type="checkbox"/> APC/HcRed	(660 nm)
<input type="checkbox"/> APC-Cy7	(785 nm)
<input type="checkbox"/> DAPI	(450 nm)
<input type="checkbox"/> AlexaFluor 430	(530 nm)

Sorting Schema:

(Please show fluorescent patterns, gating parameters and hierarchy of target cells and subsets.)



Target Subsets:

(Indicate fluorochrome positivity and negativity, e.g., FITC+ APC-, and show target subsets in the plots above.)

	~% cells in sample	~# cells needed after sort
<input type="checkbox"/> Subset 1:	_____ %	_____
<input type="checkbox"/> Subset 2:	_____ %	_____
<input type="checkbox"/> Subset 3:	_____ %	_____
<input type="checkbox"/> Subset 4:	_____ %	_____

Re-analysis of target cells after sort? yes no

Please list any special handling requests:

(e.g., shield cells from light, keep cells on ice, etc.)

For Operator Use Only:

Scheduled date and time of sort: _____

Actual start time of sort: _____ Finish time of sort: _____

Number of hours (not counting setup time):

1 hour 1-1/2 hour 2 hours 2-1/2 hours 3 hours other _____

Billing: Yes. Date of billing: _____

Result of sort:

approximate number of sorted cells (estimates only)

Subset 1 _____; Subset 2 _____; Subset 3 _____; Subset 4 _____

successful sort, no problems encountered

clumping of cells experienced

low yield

purity of sorted cells questioned

cells might be bacterially contaminated

technical problems with FACSAria

other comments:

Operator _____ Date _____